# Chitosan-platelet-rich plasma implants actions *in vitro* and *in vivo*

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PhD candidate

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ORTHO Regenerative Technologies

POLYTECHNIQUE MONTRÉAL
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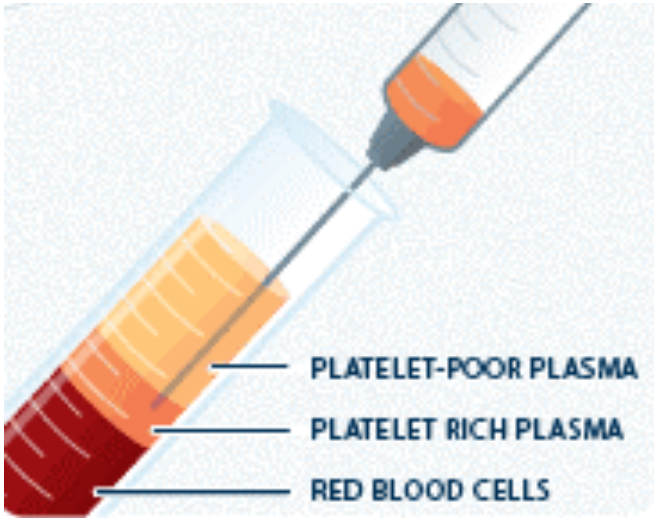
# Disclosure

- AC, NTK and MDB are shareholders and MDB is a founder of Orthoregenerative Technologies Inc

# Platelet-rich plasma

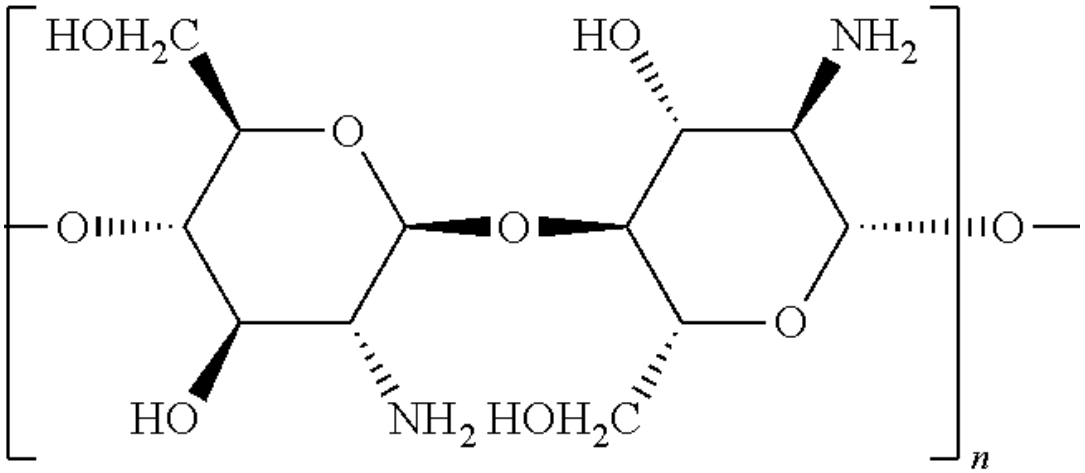
- Platelet-rich plasma (PRP), a product of whole blood is currently used in tissue regeneration.
- PRP has a high concentration of platelets and platelet-derived factors.




http://www.holisticveterinaryhealing.com/wp-content/uploads/2015/01/prp-3.png

# Chitosan

Polysaccharide obtained from partial deacetylation of chitin




chitosan

DDA= degree of deacetylation

*DDA = nD-GlcN / (nD-GlcN + nD-GlcNAc) × 100*

N-D-Glucosamine unit (GlcN)
N-acetyl-D-Glucosamine unit (GlcNAC)

# Chitosan

- High DDA chitosan (>95% DDA) is slowly degraded.
- Lower DDAs (80-85%) degrade quickly and recruit local host cells to effect tissue repair (Hoemann et al. 2010).
- Our laboratory has worked previously with chitosan-glycerol phosphate (GP)/blood implants for cartilage repair applications (BST-CarGel).
- BST-CarGel has been approved for clinical use in 17 countries.

# Previous work

- Freeze-dried Cakes contain chitosan, lyoprotectants and PRP activator, CaCl2

- Freeze-dried Cakes dissolve readily in PRP and coagulate rapidly to make homogenous chitosan-PRP hybrids that

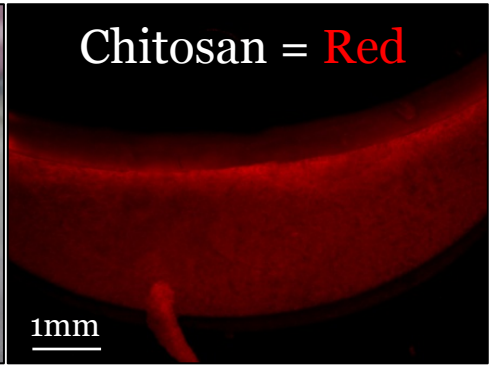
1) do not shrink, versus up to 90% volume loss in PRP

2) produce sustained biological activity

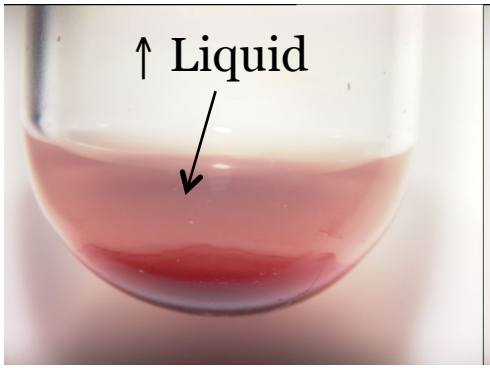
3) have in situ tissue building capacity

Freeze-dried cake

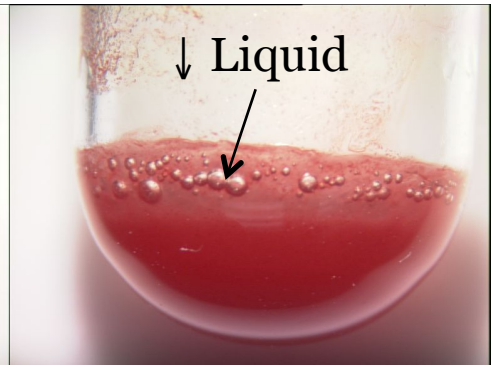

Hybrid clot




PRP only clot




Hybrid clot




*(Chevrier et al BMM 2016 submitted)*

# Objectives

- Objective 1→ investigate chitosan-PRP hybrid clot retraction
- Objective 2→ investigate the levels of growth factors released from chitosan-PRP in culture medium.
- Objective 3 → characterize the effect of chitosan and lyoprotectant on platelet activation *in vitro*
- Objective 4 → inject freeze-dried chitosan/PRP implants subcutaneously in rabbit to assess the effect of DDA on cell recruitment.

# Objective 1:
# Clot retraction

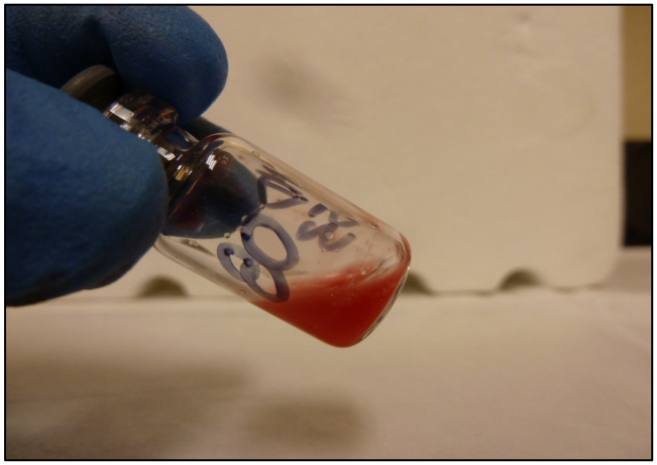
- Hypotheses:

- Chitosan binds to platelets →inhibits platelet aggregation which is required for strong clot retraction.

- The fibrin network formed in the presence of chitosan in chitosan-PRP hybrid clots is similar to the network formed PRP-only clots.

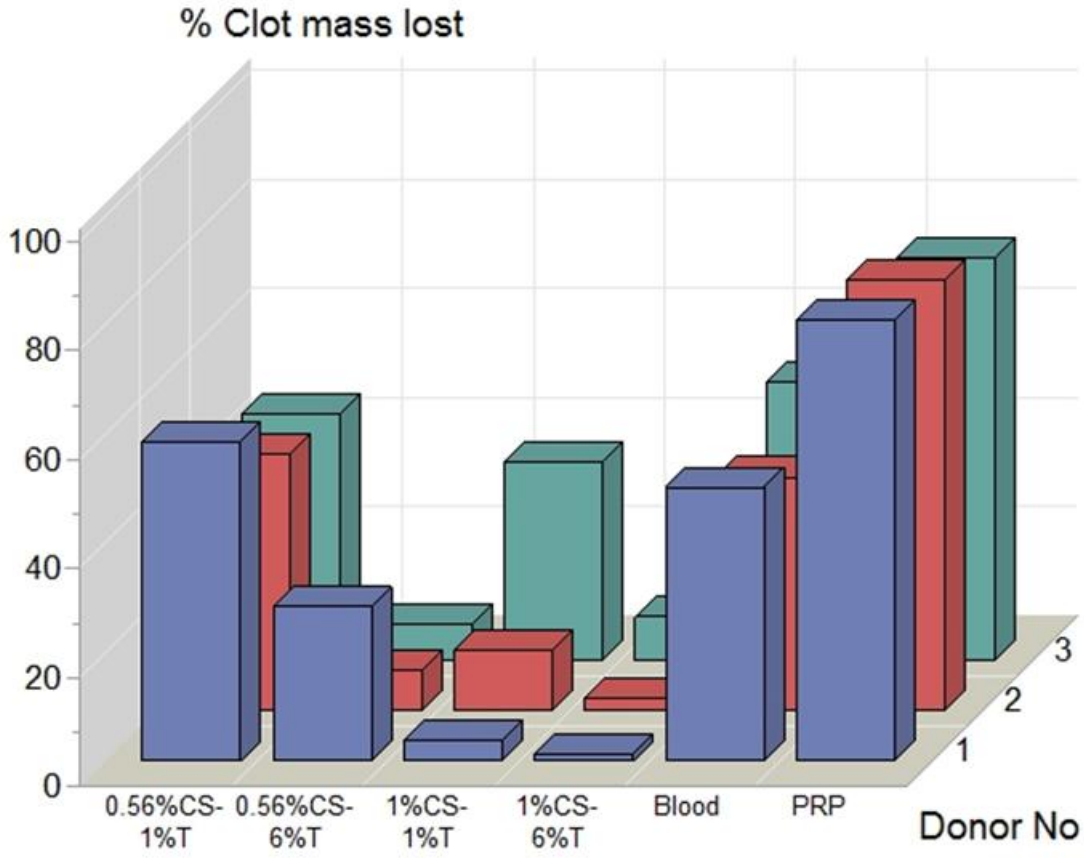
# Clot retraction

- Methods:
- Four freeze-dried formulations solubilized in PRP
- Gravimetric measurements and Imaging on clots



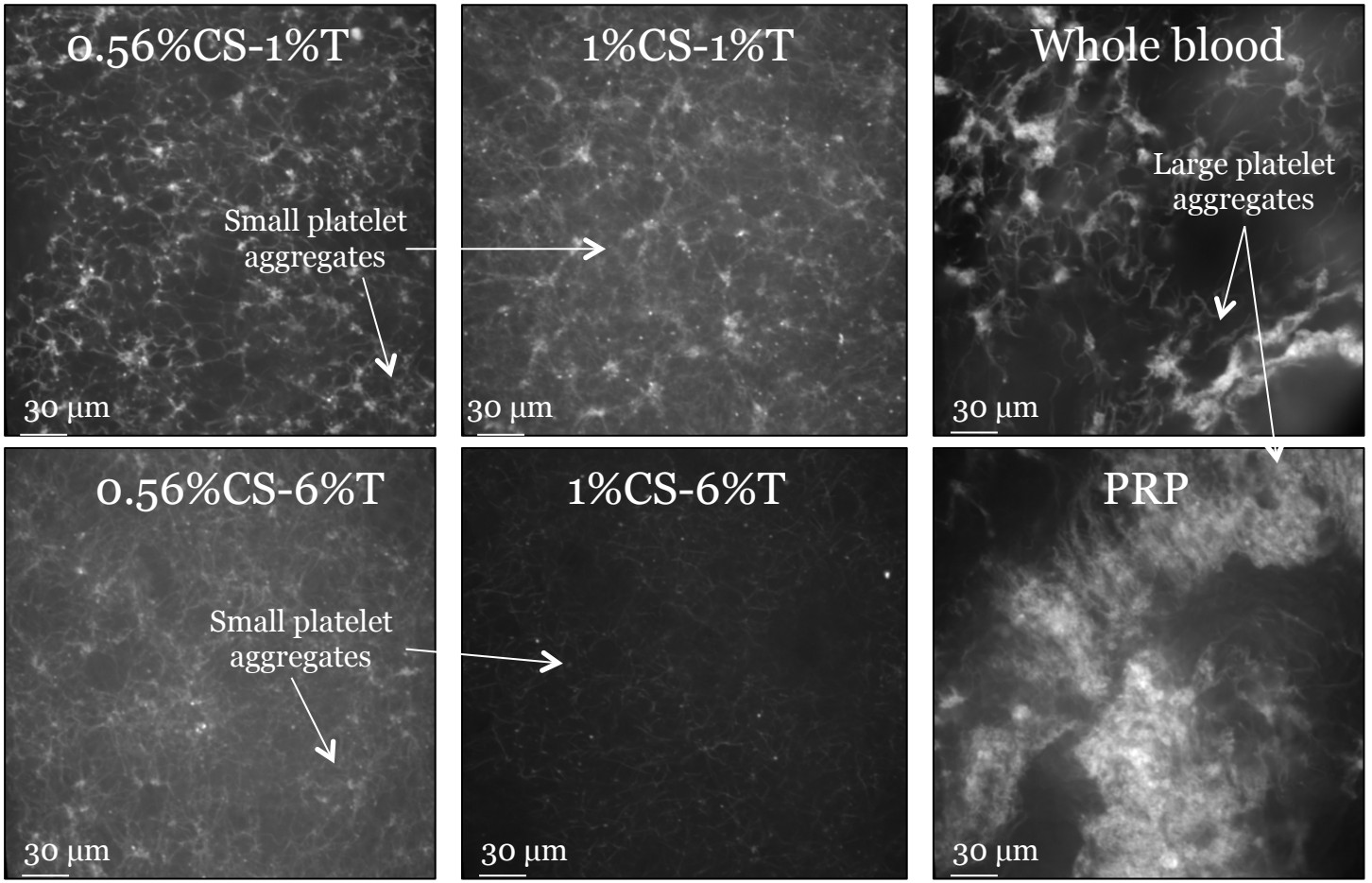
| CS | Sol | Formulation | Chitosan (w/vol) | HCl for 60% protonated (mM) | Trehalose (mM) | $CaCl_2$ (mM) | Aliquot into | Rehydrated in (volume) of PRP |
|---|---|---|---|---|---|---|---|---|
| | 1 | 0.56% CS-1% Trehalose | 0.56% | 16mM | 26mM | 42.2mM | 1mL | 1mL |
| | 2 | 0.56% CS-6% Trehalose | 0.56% | 16mM | 159mM- | 42.2mM | 1mL | 1mL |
| | 3 | 1% CS-1% Trehalose | 1% | 29mM | 26mM | 42.2mM | 1mL | 1mL |
| | 4 | 1% CS-6% Trehalose | 1% | 29mM | 159mM- | 42.2mM | 1mL | 1mL |

# Chitosan-PRP hybrid clots inhibit retraction

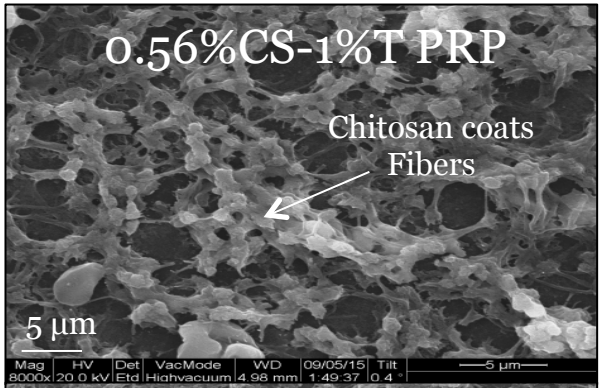


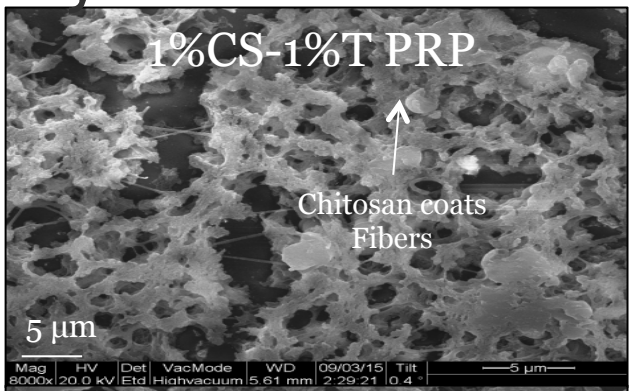

% Clot mass lost after clotting for 1 hour at 37°C for donor 1 (blue bars), donor 2 (red bars) and donor 3 (green bars) at 37°C. n = 2 clots for each measurement, with bars showing average of 2 clots.

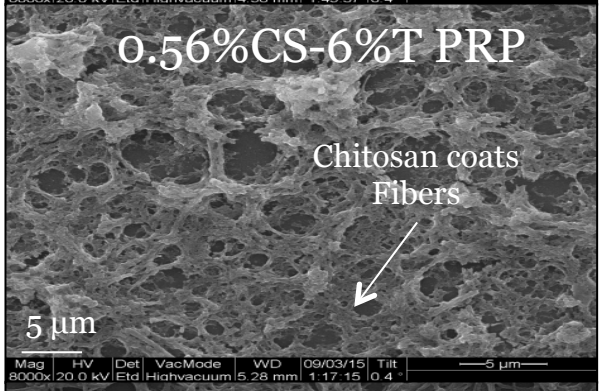
# Platelet aggregates are smaller in chitosan-PRP hybrid clots and fibrin network is finer in presence of high trehalose

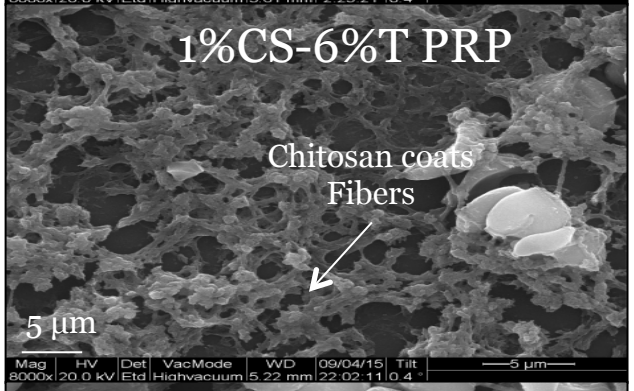


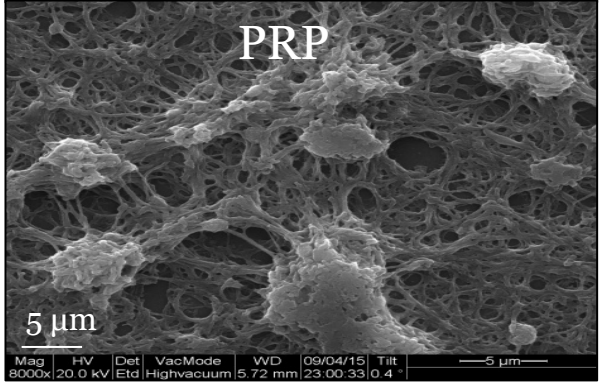
# Cells and fibers are covered with chitosan in chitosan-PRP hybrid clots

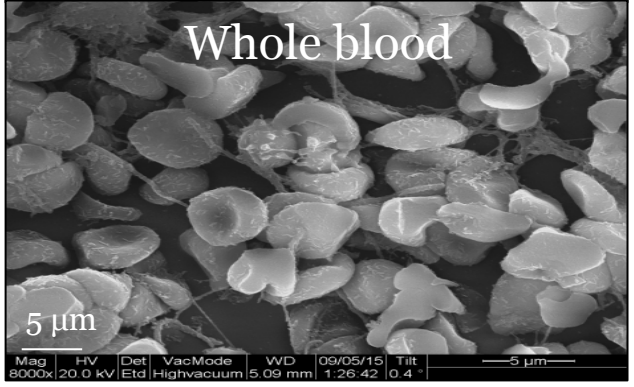


# Conclusion: Objective 1

- Whole blood clots and PRP clots retracted significantly upon coagulation.
- Clot retraction was inhibited in chitosan-PRP hybrids.

# Objective 2:
# Growth Factor Release Profiles

Methods:

- One formulation (1% CS-1% Trehalose) solubilized in PRP from three donors → Duplicate clots from each donor cultured for 7 days, followed by ELISA for PDGF-AB, EGF and VEGF.

# Release profiles

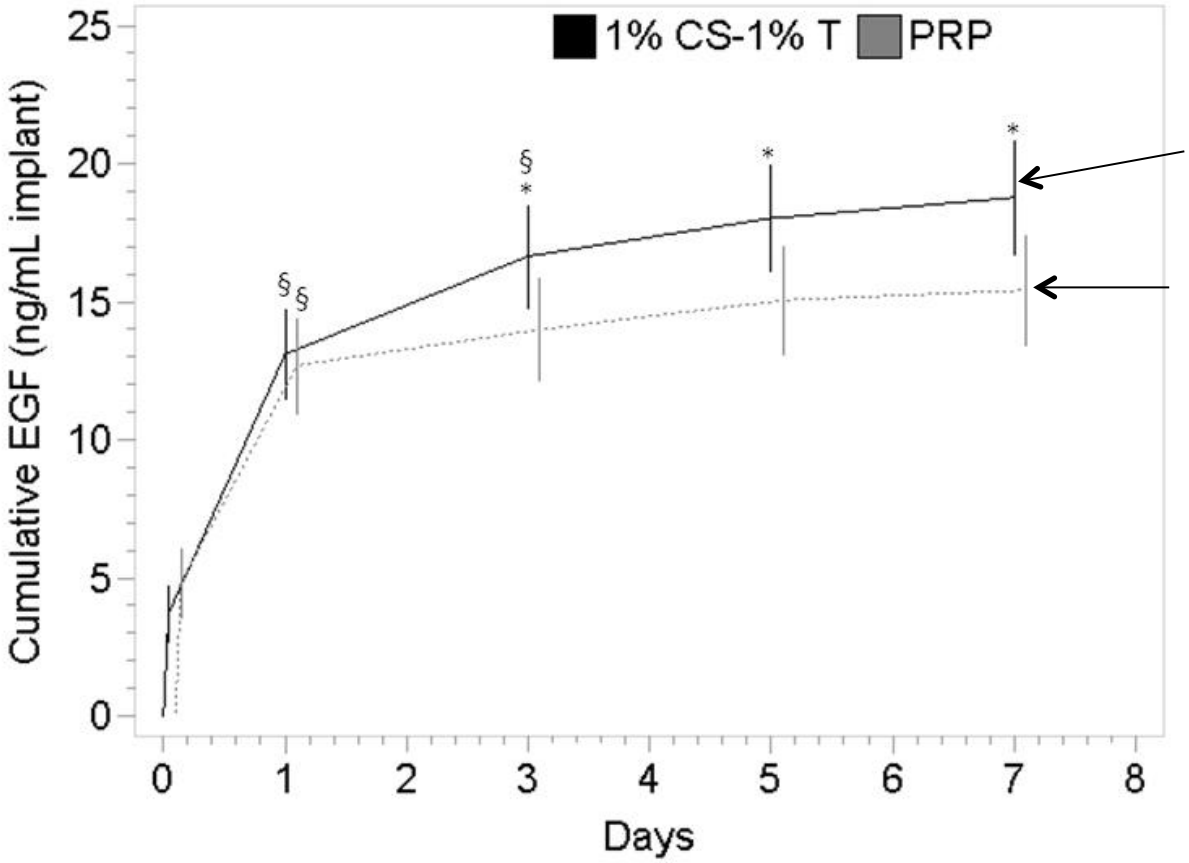
- Hypotheses:

- Negatively charged growth factors with low isoelectric points, would bind to positive CS to slow release.

- Positively charged growth factors with high isoelectric points would burst release,  due to ionic repulsion.

| Growth factors | PDGF-AB | VEGF | EGF |
|---|---|---|---|
| Isoelectric point | 9.8 | 8.5 | 4.6 |
| Charge | Positive | Positive | Negative |

# Continuous EGF release from hybrid and control clots and higher cumulative release from CS-PRP



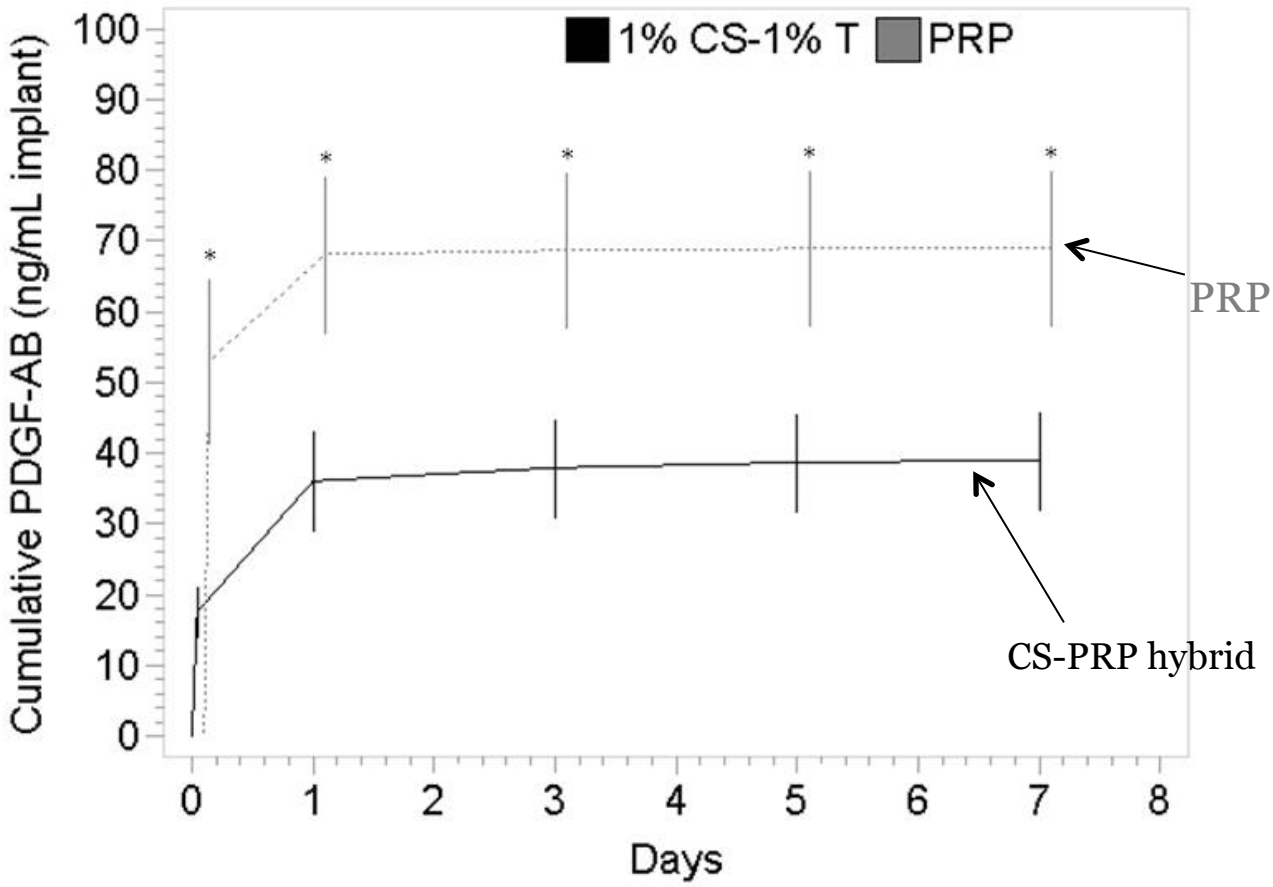

CS-PRP hybrid

PRP

-Implant 250µL
-Culture medium 500µL
-Medium collected at 0, 1, 3, 5, & 7 days

Results are presented as Mean ± SE. n = 6 clots. § $p < 0.05$ for time point immediately prior.

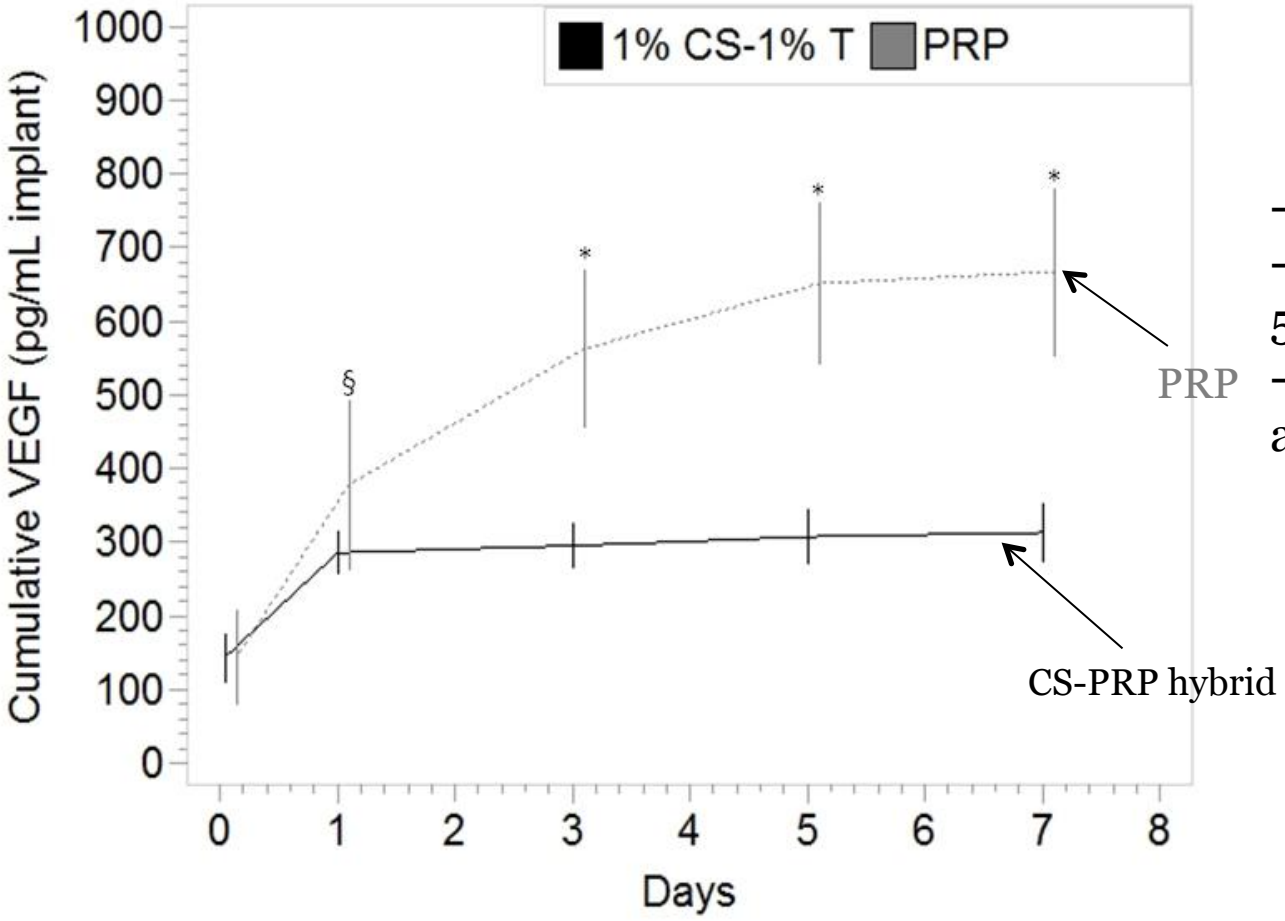
# Burst PDGF-AB release from hybrid and control clots and lower cumulative release from CS-PRP




-Implant 250µL
-Culture medium 500µL
-Medium collected at 0, 1, 3, 5, & 7 days

Results are presented as Mean ± SE n = 6 clots. § $p < 0.05$ for time point immediately prior.

# Burst VEGF release from hybrid clots and continuous release from control clots




-Implant 250µL
-Culture medium 500µL
-Medium collected at 0, 1, 3, 5, & 7 days

Results are presented as Mean ± SE. n = 6 clots. § $p < 0.05$ for time point immediately prior.

# Conclusions of Objective 2

- High inter-individual variation between the amount of growth factor released by each donor, but the patterns of release were similar for all three donors.

- Release profiles did not support our charge-dependent hypotheses. Investigations into how growth factors bind to chitosan in our hybrids are still ongoing .

# Objective 3:
# Characterize the effect of chitosan and lyoprotectant on platelet activation *in vitro*

Hypothesis:

- Chitosan and trehalose will induce platelet activation.

# Platelet activation

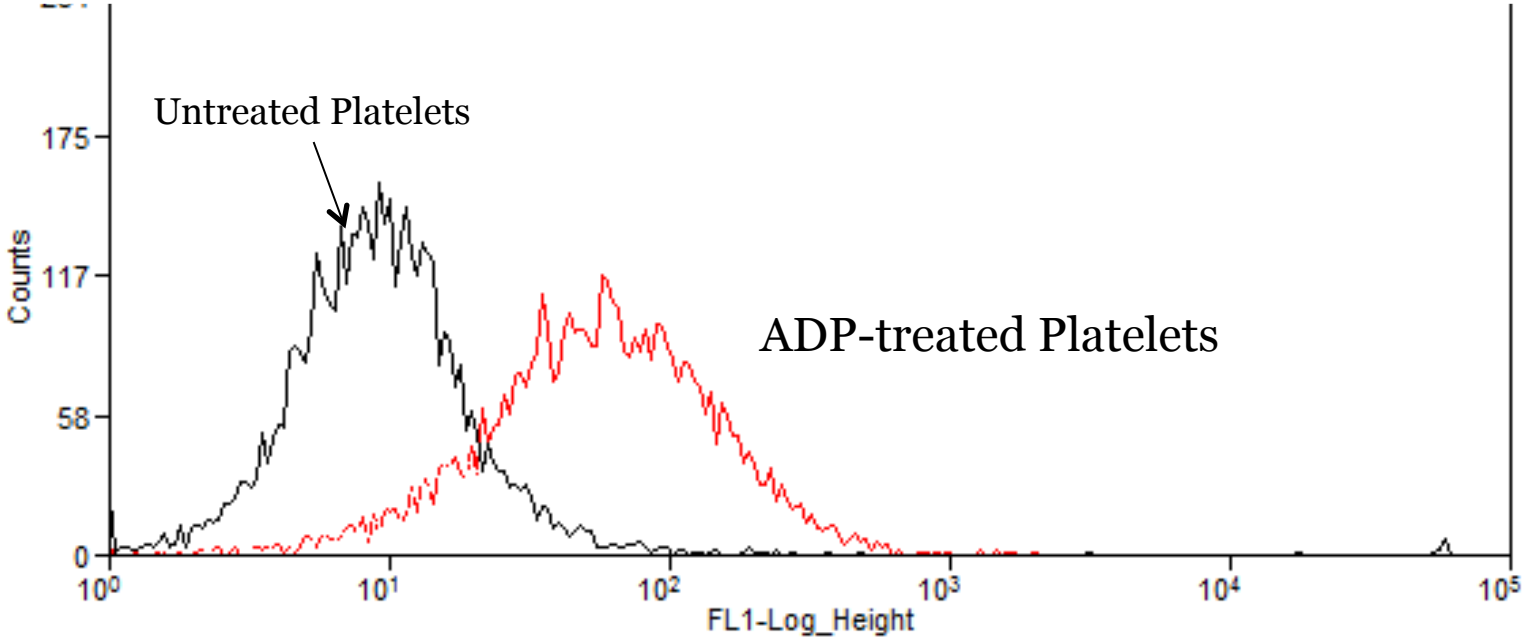
Methods proposed:

- Isolated platelets flow cytometry → assess platelet function after contact with chitosan and/or lyoprotectant.

- Monoclonal antibody :
  - Pac-1 fluorescein-conjugated antibodies (Activated αIIbβ3).

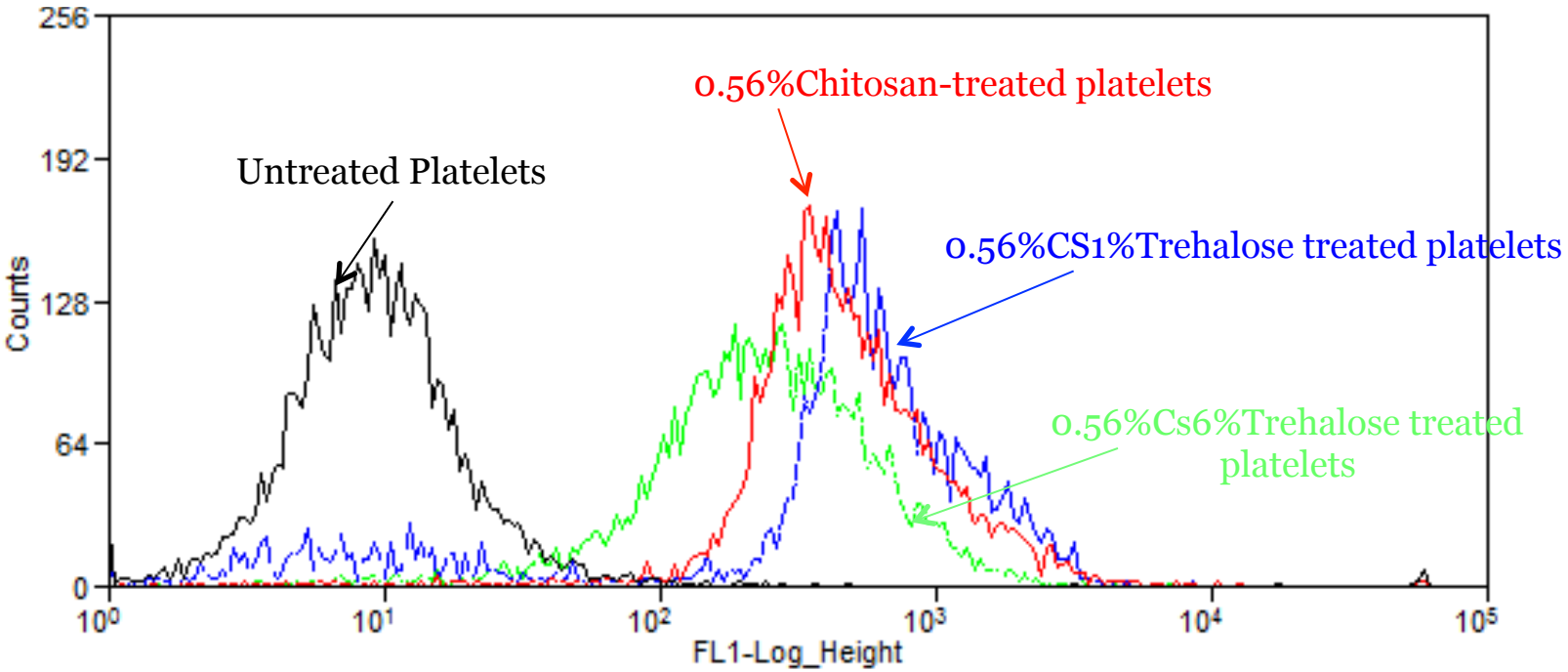
# Platelet activation

## PAC-1 Flow cytometry

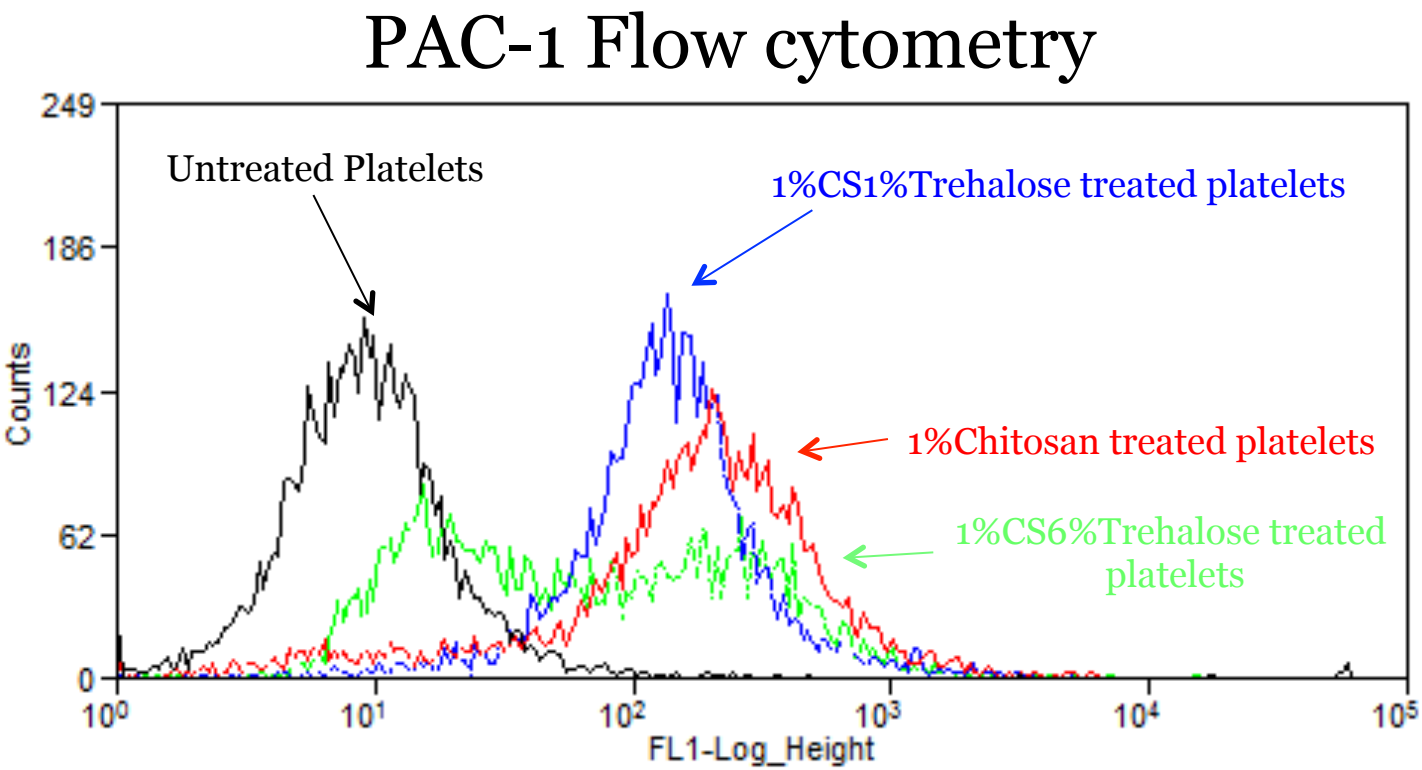


# Platelets activated in contact with 0.56% chitosan

## PAC-1 Flow cytometry

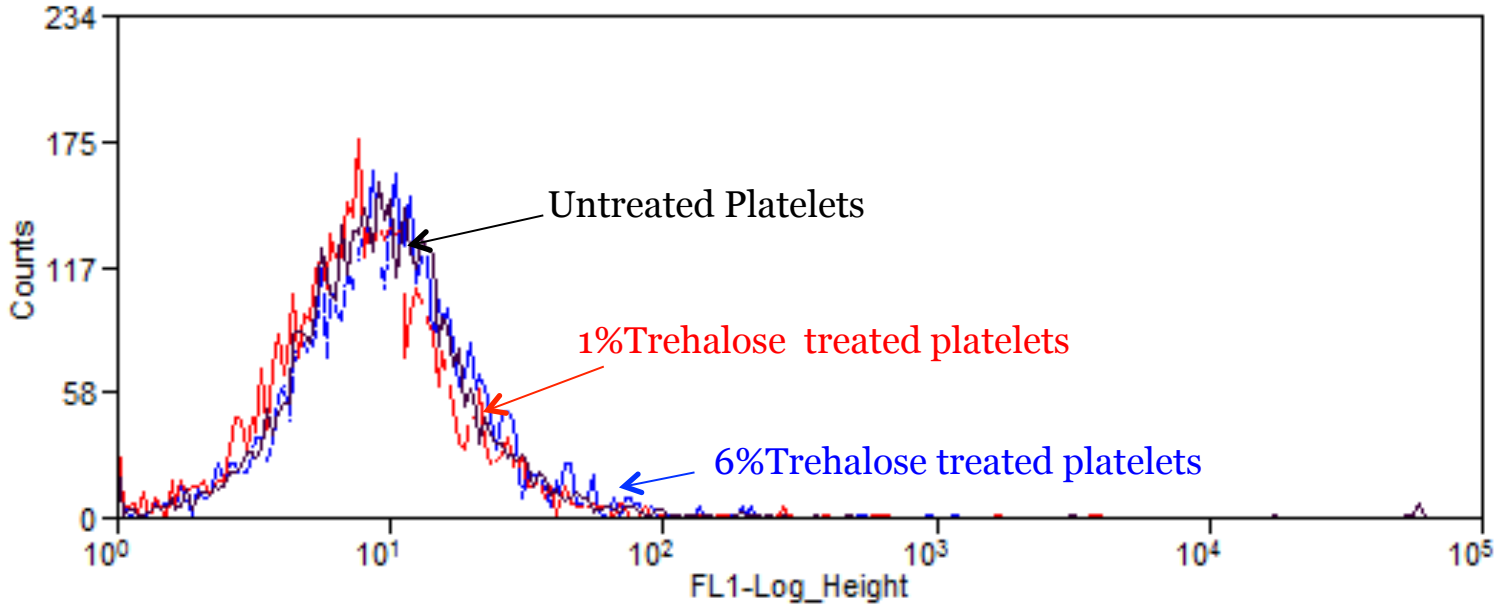


# Platelets activated in contact with 1% chitosan



# Lyoprotectant alone does not activate platelets

## PAC-1 Flow cytometry



# Conclusions of Objective 3

- Platelets are activated when placed in contact with chitosan (0.56% or 1%) with or without lyoprotectant.
- No activation with lyoprotectant only.
- Inhibition of clot retraction is not due to inhibited platelet activation.

# Objective 4
# Subcutaneous implants

- Methods:

- 4 different Chitosan/PRP formulations ($M_n$ *40kDa 1% (w/v) CS, 1% (w/v) trehalose and DDA 80%, 85%,90%, 95%*) were injected subcutaneously in the back of five rabbits.

| **Test Article** | **Total Volume (mL)** | **# Animals / time points** | | |
|---|---|---|---|---|
| Chitosan-PRP | 150 µL per implant | n = 2 at day 14 | n = 2 at day 28 | n = 1 at day 42 |

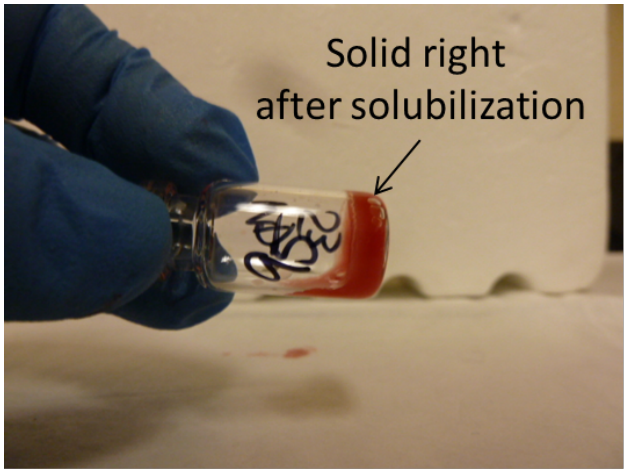
# Subcutaneous implants

- Hypotheses:

- Chitosan-PRP formulations containing chitosan of higher DDA will reside longer subcutaneously *in vivo*.

- Chitosan-PRP formulations containing chitosan of lower DDA will induce a greater acute inflammatory reaction (Lafantaisie-Favreau, C.-H., Hoemann, C. D. 2013).

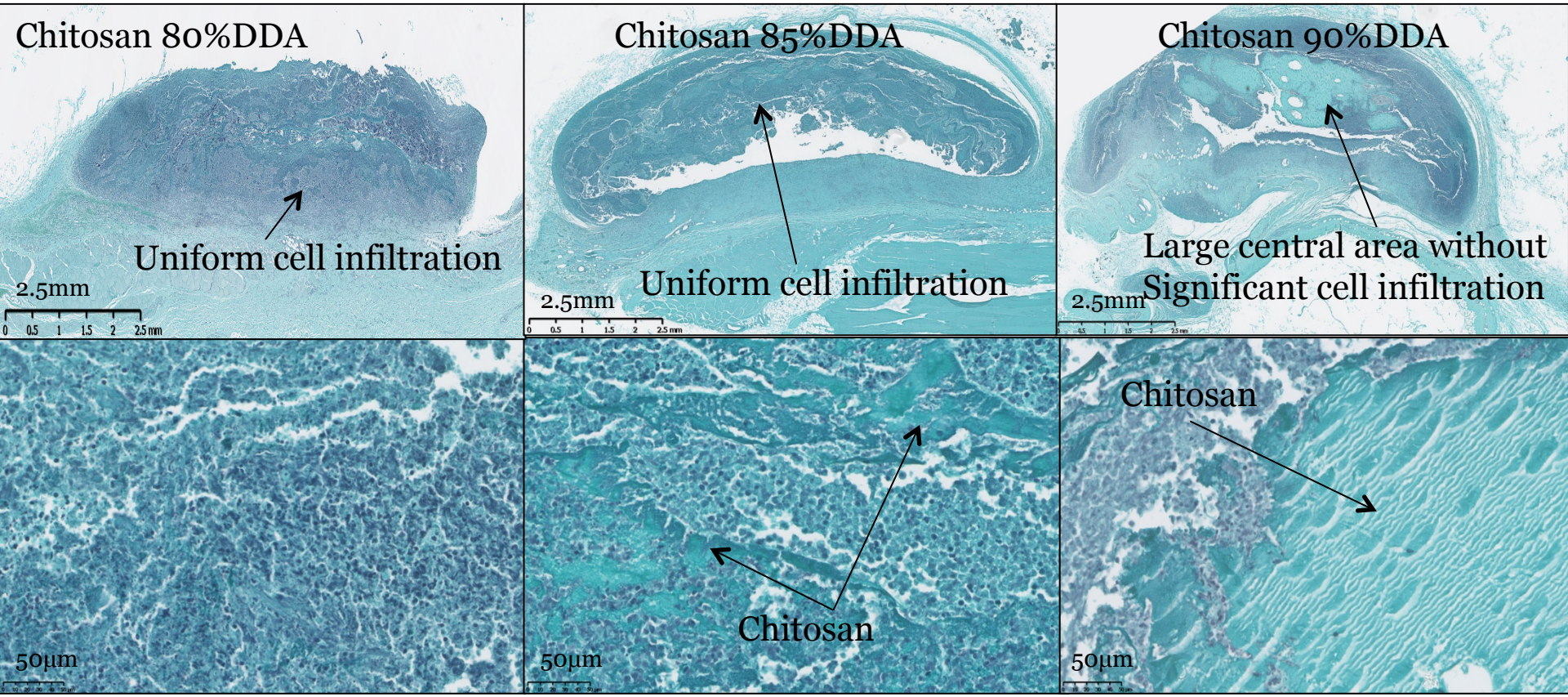
# 95% DDA chitosan-PRP solidified immediately





Erythrocyte agglutination induced by chitosan?

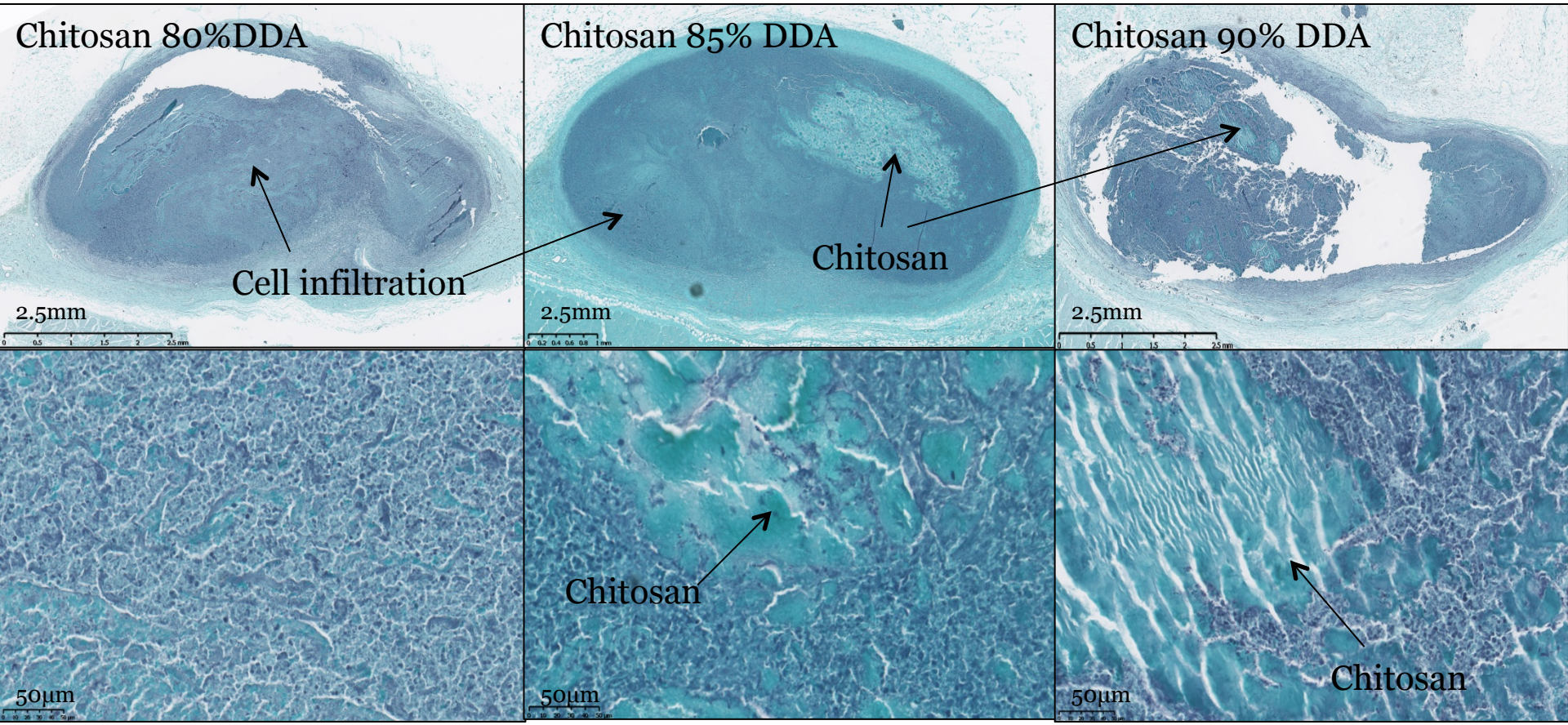
# Cell infiltration is more uniform with chitosan of lower DDA at 2 weeks




Iron hematoxylin/Fast Green stained paraffin section of chitosan-PRP
Bottom pictures magnitude 40X

*PRP alone completely degraded at two weeks

# Chitosan-PRP implants persist until 6 weeks in this model




Iron hematoxylin/Fast Green stained paraffin section of chitosan-PRP
Bottom pictures magnitude 40X

# Conclusions of Objective 4

- Chitosan-PRP implants were resident until at least 6 weeks post-implantation → prolonged bioactivity *in vivo*.
- Invasion of the implants by host cells → influenced by the degree of deacetylation of the chitosan at early time points.
- Size of the implants decreased with time.

# Overall Conclusions

- Platelet aggregation and clot retraction are inhibited in chitosan-PRP hybrid clots; platelets are activated.
- Residency of chitosan-PRP in vivo leads to increased biological activity in vivo compared to PRP alone.
- We are currently testing the efficacy of Chitosan-PRP in Rotator cuff tear repair in rabbit and sheep models.

# Acknowledgements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